

## Chemically Competent Cells

**BE SURE TO PERFORM THIS IN A COLD, STERILE ENVIRONMENT. READ AND PREPARE BEFORE PERFORMING.**

- Grow overnight culture of desired strain you want to make Chemically Competent. This is your seed stock.
- Inoculate 10 ml of SOB medium with seed stock and grow at 37°C to an OD<sub>600nm</sub> of  $\sim 0.5$  <sup>or LB</sup>  $\downarrow$   $0.05$
- Place inoculum on ice for 30 minutes
- Shake frequently to maintain homogeneous solution
- Centrifuge at 4000rpm at 4°C for 10 minutes.
- Discard supernatant by pouring out slowly and pipetting gently or mix by “flicking” remaining supernatant.
- Gently re-suspend in 10 ml of ice cold CCMB80 buffer.
- Incubate on ice 20 minutes
- Centrifuge again at 4°C and discard supernatant as described above.
- Re-suspend in ice cold CCMB80 buffer to an OD of  $\sim 3.0$
- Aliquot into 2.0 mL graduated tubes
- Label appropriately and store at -80°C.

Repeat

## Heat Shock Transformation SOP

1. Pre-chill 15mL sterile **culture tube** on ice.
2. Place Competent Cells on wet ice until just thawed.
3. Gently transfer 50  $\mu$ l of Competent Cells to pre-chilled 15mL culture tube.
4. Add 1-2  $\mu$ l of Ligated DNA (either Registry BioBricks from distribution kit or ligated parts). Move pipette tip through the cells while dispensing. Gently (VERY GENTLY) tap <sup>DN A</sup> the tube once or twice.   
min. 100ms
5. Immediately return tube to ice for 30 min.
6. Place tube in 42°C water bath for 45-50 sec (DO NOT SHAKE).
7. Immediately place the tube on ice for 2 min.

or LB + glucose  
↳ 100  $\mu$ L  
of 200 g/L  
solution  
60-120

8. Add 950  $\mu$ l of cold SOC medium and incubate for ~~60-90~~ minutes at 37°C with shaking.
9. Centrifuge tubes @4000rpm for 3 minutes and discard supernatant. Resuspend cells in remaining solution and use 50 $\mu$ L to spread onto LB with appropriate antibiotic.
10. Incubate the plates overnight at 37°C.