

# Annealing Oligos

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## Introduction

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## Materials

- › Nuclease-free TE
  - › We resuspend in TE to help suppress nuclease activity that would degrade the primers.
- › 1M NaCl solution (in water)
- › Dehydrated oligos from IDT
- › One thin-walled 200  $\mu$ l PCR tube
- › One epi tube (required), plus another (optional)

## Procedure

### Preparing oligos

1. Resuspend the oligos at 100  $\mu$ M, as described in "Resuspending Primers"
2. Set up a thermocycler program for the following annealing program:
  - Heat to 95° for 2 minutes
  - Every 1'30", decrease the temperature by 1° to a final temperature of 25°
  - Hold at 4°

### Procedure

3. In the thin-walled PCR tube, mix:
  - 75  $\mu$ l nuclease-free TE
  - 5  $\mu$ l 1M NaCl
  - 10  $\mu$ l oligo #1
  - 10  $\mu$ l oligo #2
  - (This is 10 $\mu$ M)
4. Place tube in the thermocycler and run the annealing program
5. Transfer annealed oligo to the epi tube. Label with name and concentration (10  $\mu$ M) and store at -20°C.
6. For setting up Golden Gate reactions: 1  $\mu$ M is 1000 fmol/ $\mu$ l. If your desired input to a GoldenGate reaction is 50 fmol, then:
  - Add 199  $\mu$ l of TE to the second epi tube
  - Add 1  $\mu$ l of the 10  $\mu$ M duplex

- Mix well, label with name and concentration (50 fmol/ $\mu$ l) and store at -20°.