

## **Media & Antibiotics**

(by Karen, 2010, updated Julia 2014)

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## Media

### Luria-Bertani (LB) broth:

Tryptone	10 g
Yeast extract	5 g
NaCl	10 g
H <sub>2</sub> O (dest)	ad 1.000 ml

- for LB plates: add 15 g/l of agar
  - important: cool down the agar solution to 50°C before adding antibiotics

### SOB medium

5 g	NaCl
20 g	tryptone
5 g	yeast extract
2.5 ml	1M KCl
Add water to 1L and autoclave.	

### Difco Sporulation Medium (DSM):

Nutrient Broth	8 g
KCl	1 g
MgSO <sub>4</sub> (1 M)	1 ml
MnCl <sub>2</sub> (10 mM)	1 ml
H <sub>2</sub> O (bidest)	ad 1.000 ml
Add after autoclave:	
CaCl <sub>2</sub> (1 M)	0,5 ml (500 µM end)
FeSO <sub>4</sub> (1 mM)	1 ml (1 mM end)

- for DSM plates: add 15 g/l of agar
  - important: cool down the agar solution to 50°C before adding antibiotics

## **Starch plates:**

Nutrient Broth (Difco)	7,5 g
Starch	5 g
Agar	15 g
H <sub>2</sub> O (dest)	ad 1.000 ml

## **Chemical defined medium (CSE):**

5×C-Salts	20 ml
Tryptophan (5 mg/ml)	1 ml
Ammoniumeisencitrat (2,2 mg/ml)	1 ml
III'-Salts	1 ml
Potassium glutamate (40%)	2 ml
Sodium succinate (30%)	2 ml
Sterile H <sub>2</sub> O	to 100 ml

### 5×C-Salts (1 l)

KH <sub>2</sub> PO <sub>4</sub>	20 g
K <sub>2</sub> HPO <sub>4</sub> × 3 H <sub>2</sub> O	80 g
(NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	16,5 g

### III'-Salts (1 l)

MnSO <sub>4</sub> × 4 H <sub>2</sub> O	0,232 g
MgSO <sub>4</sub> × 7 H <sub>2</sub> O	12,3 g

- autoclave (or filtrate) each component separately and put them together freshly before starting your experiment
- For strains carrying an integration in *thrC* also add 1 ml per 100 ml threonine (5 mg/ml)
- Optional: addition of media additives, for example pyruvate (0.5% final concentration), glucose (1% final concentration) or other C-source. (Optimal xylose-regulation and extended exponential growth phase is achieved with 2.5% fructose as C-source)

## **Modified chemically defined medium (MCSE)**

10×MOPS solution	10 ml
Tryptophan (5 mg/ml)	1 ml
Ammoniumeisencitrat (2,2 mg/ml)	1 ml
III'-Salts	1 ml
Potassium glutamate (40%)	2 ml
Sodium succinate (30%)	2 ml
Fructose (20%)	1 ml
Sterile H <sub>2</sub> O	to 100 ml

### **10 x MOPS solution (1 l) adjust pH 7 with KOH (10 M)**

**(=400 mM MOPS, 10 mM Phosphate)**

MOPS	83,72 g
(NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	33g
KH <sub>2</sub> PO <sub>4</sub> (1M)	3,85 ml
K <sub>2</sub> HPO <sub>4</sub> (1M)	6,15 ml

### **III'-Salts (1 l)**

MnSO <sub>4</sub> × 4 H <sub>2</sub> O	0,232 g
MgSO <sub>4</sub> × 7 H <sub>2</sub> O	12,3 g

- autoclave (or filtrate) each component separately and put them together freshly before starting your experiment
- For strains carrying an integration in *thrC* also add 1 ml per 100 ml threonine (5 mg/ml)

## Soft agar:

Tryptone	10 g
Yeast extract	5 g
NaCl	10 g
Agar	7.5 g (0.75%)
H <sub>2</sub> O (dest)	ad 1.000 ml

## Mueller Hinton Medium:

Mueller-Hinton Broth	21 g
H <sub>2</sub> O (dest)	ad 1.000 ml

## 2 × YT:

Tryptone	16 g
Yeast extract	10 g
NaCl	5 g
H <sub>2</sub> O (dest)	ad 1.000 ml

## Thr + / Thr- MNGE Plates (for ThrC insertion test)

Part1: 3% Agar (125 ml H<sub>2</sub>O + 3,75 g Agar) -> autoclave

Part2: 2 x MNGE:	10 ml	10x MN (autoclaved)
	92 ml	dH <sub>2</sub> O (autoclaved)
	25 ml	Glucose (20 %) (sterile filtered)
	1,25 ml	Kalium-Glutamate (40 %) (sterile filtered)
	1,25 ml	Ammonium-Fe-Citrat (2,2 mg/ml) (autoclaved)
	2,5 ml	Tryptophan (5 mg/ml) (sterile filtered)
	250 µl	MgSO <sub>4</sub> (1M) (sterile filtered)

Thr + plates: add 2,5 ml threonine (5 mg/ml) (sterile filtered)

Thr – plates: add nothing more

Mix part1 and part2 at appropriate temperature (handwarm) and pour plates.

**Sistrom's Minimal Medium (SIS) for *Rhodobactersphaeroides* (Sistrom, 1962):**

**10x Medium**

	<b>Component</b>	<b>Chemical</b>	<b>MW</b>	<b>For 1L</b>
<b>or</b>	Dipotassiumhydrogenphosphate anhydrous	K <sub>2</sub> HPO <sub>4</sub>	174.18	34.8 g
	Potassium dihydrogen phosphate	KH <sub>2</sub> PO <sub>4</sub>	136.09	27.2 g
<b>or</b>	Ammonium sulfate	(NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	132.14	5 g
	Ammonium chloride	NH <sub>4</sub> Cl	53.49	1.95 g
	Succinic acid	C <sub>4</sub> H <sub>6</sub> O <sub>4</sub>	118.09	40 g
<b>or</b>	L-Glutamic acid potassium salt monohydrate	C <sub>5</sub> H <sub>8</sub> NO <sub>4</sub> K*H <sub>2</sub> O	203.23	1.38 g
	L-Glutamic acid	C <sub>5</sub> H <sub>9</sub> NO <sub>4</sub>	147.13	1 g
	L-Aspartic acid	C <sub>4</sub> H <sub>7</sub> NO <sub>4</sub>	133.1	0.4 g
	Sodium chloride	NaCl	58.44	5 g
	Nitrilotriacetic acid	C <sub>6</sub> H <sub>9</sub> NO <sub>6</sub>	191.14	2 g
	Magnesium sulfate	MgSO <sub>4</sub> *7H <sub>2</sub> O	246.48	3 g
<b>or</b>	Calcium chloride dihydrate	CaCl <sub>2</sub> *2H <sub>2</sub> O	147.02	0.334 g
	Calcium chloride	CaCl <sub>2</sub>	110.98	0.250 g
	Ferrous sulfate	FeSO <sub>4</sub> +7H <sub>2</sub> O	278.02	0.02 g
	1% Ammonium molybdate	(NH <sub>4</sub> ) <sub>6</sub> Mo <sub>7</sub> O <sub>24</sub> *4H <sub>2</sub> O	1235.86	0.2 ml
	Trace element solution			1 ml
	Vitamin solution			1 ml
Dissolve in 1L H <sub>2</sub> O, aliquot in bottles and store at -20°C				

**Trace element solution**

EDTA	1.765 g
ZnSO <sub>4</sub> *7H <sub>2</sub> O	10.950 g
FeSO <sub>4</sub> *7H <sub>2</sub> O	5 g
MnSO <sub>4</sub> *H <sub>2</sub> O	1.54
CuSO <sub>4</sub> *5H <sub>2</sub> O	0.392 g
Co(NO <sub>3</sub> ) <sub>2</sub> *6H <sub>2</sub> O	0.248 g
H <sub>3</sub> BO <sub>3</sub>	0.114 g
Dissolve in 100 ml H <sub>2</sub> O, store at 4°C	

### Vitamin solution

Nicotinic acid	1 g
Thiamine-HCL	0.5 g
Biotin	0.01 g
Dissolve in 100 ml H <sub>2</sub> O, store at 4°C	

- prepare 1x medium
- adjust pH to 7.0 with KOH (for 1L medium ≈ 3.5 g KOH pellets)
- for plates, add agar to 15 g/L
- autoclave

## Antibiotics

- Indicated are 1,000-times stock solutions
- Dissolve in the specific solvent and filtrate by using 0.2 µm filters
- Store at -20°C

Strain	Antibiotic	Concentration	Dissolve in	Color code
<i>B. subtilis</i>	Kanamycin	10 mg/ml	H <sub>2</sub> O	Black (one bar)
	Chloramphenicol	5 mg/ml	70% ethanol	Blue (one bar)
	MLS selection:			Red
	Erythromycin	1mg/ml	70% ethanol	
	Lincomycin	25 mg/ml	H <sub>2</sub> O	
	Spectinomycin	100 mg/ml	H <sub>2</sub> O	Purple
	Tetracyclin <sup>1</sup>	12,5mg/ml	--	Orange
<i>E. coli</i>	Bacitracin	50 mg/ml	H <sub>2</sub> O	-
	Daptomycin	2 mg/ml	H <sub>2</sub> O	-
	Ampicillin	100 mg/ml	H <sub>2</sub> O	Green
	Chloramphenicol	35 mg/ml	70% ethanol	Blue (two bars)
<i>Rhodobacter</i>	Kanamycin	50 mg/ml	H <sub>2</sub> O	Black (two bars)
	Streptomycin	20 mg/ml		?
	Tetracyclin	1 mg/ml	H <sub>2</sub> O	Orange
<i>Streptomyces</i>	Spectinomycin	25 mg/ml	H <sub>2</sub> O	Purple
	Kanamycin	25 mg/ml	H <sub>2</sub> O	Black
	Apramycin	50 mg/ml	H <sub>2</sub> O	Pink
	Thiostrepton	50 mg/ml	DMSO	-
	Nalidixic Acid	25 mg/ml	H <sub>2</sub> O	-

<sup>1</sup> Ordered from Bioline, already in solution (90 % EtOH)

## Further substances

### IPTG, X-Gal, BCIP

- IPTG: 1 M (dissolve in H<sub>2</sub>O; store at -20°C)
- X-Gal: 100 mg/ml (dissolve in DMF; color code: blue X; store at -20°C)
- BCIP (5-Brom-4-chlor-3-indoxylphosphat-p-tolidinsalz): 50 mg/ml (dissolve in DMF; store at -20°C)

### Iod-Potassiumiodine solution (Starch- und Ozondetektion: blue)

Solve 2g Pottassiumiodine in 10 ml H<sub>2</sub>O and add 1 g Iod (resublimiert) (under fume hood). Fill to 100 ml with H<sub>2</sub>O. Use Braun Glasflask (or wrap with Alu-folie foil).

### Bacitracin stock-solution (by Susanne, 2011)

#### Zinc salt (e.g. Sigma B-5150) for Induction (Stock solution 20 mg/ml)

- Soluble in water at acidic pH up to ca. 20 mg/ml
- Weigh desired amount into an eppendorf tube
- Add correct amount of dH<sub>2</sub>O (minus ca. 5 µl) and vortex to get a homogenous suspension
- Add 10% HCl 1 µl at a time, mix and check if solution clears (add more acid if not)
- Solution is stable for a long time at -20°C and can take many rounds of thawing and re-freezing

Note: For 1 ml @ 10 mg/ml I normally need around 2-3 µl acid (20 mg/ml solutions take much more)

#### Zinc-free Bacitracin

- Easily soluble in water, up to 100 mg/ml possible
- No acid needed

Note: For biological activity, Bacitracin needs the Zn<sup>2+</sup> ion, so if you work with Zinc-free bacitracin you have to make sure there's Zinc in the media or there will be no antimicrobial activity!!!