

Lab Notebook - Week 11 (8/21/2017-8/27/2017)

Project: NU iGEM 2017 Shared Project

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MONDAY, 8/21/17

Gibson PCR product concentrations

NapA insert 15.7 ng/uL - dilute to 20nM

NapA bb 96.3 ng/uL - dilute to 20nM

YcdB bb 83.7 ng/uL -dilute to 20nM

YcdB ins 18.9 ng/uL -dilute to 20nM

Gibson YcdB

10uL of Gibson MM

1uL of YcdB bb (20nM)

3uL of YcdB ins (20nM)

6uL nf water

Gibson Transformation: YcdB

- Thaw comp cells (50uL) on ice for 15 minuts
- Pipette 15uL of Gibson assembly DNA into comp cells
- Incubate on ice for 20min.
- Heat shock at 42°C for 60s
- Incubate on ice for 5min.
- Add 65uL of SOC medai, rescue for 1:15 min in shaker at 37°C
- Plate 80uL onto Cm plate and put in incubator at 37°C for 18 hours.

SWAP cjCAS9 and saCAS9

YcbK-cjCas9-His6

PCR backbone:

Primers: P81 (FWD)

P95 (REV)

DNA: YcbK-saCas9-His6

resuspend primers to 100uM

dilute primers from 100uM to 10uM

2.5uL of 100uM P81/P95

22.5uL of nf water each

PCR components:

nf water: 22uL

P81: 1uL

P95: 1uL

DNA Plasmid: 1uL

Phusion HS MM: 25uL

*Dilute YcbK-saCas9-His6 to 10ng/uL

PCR Protocol:

98°C - 30s

10 cycles

98°C - 15s

57.4°C - 30s

72°C - 1:09 min

25 cycles

98°C - 15s

69.5°C - 30s

72°C - 1:09 min

72°C - 5min

4°C - inf.

Test NapA primers

Primers: P89, P67

DNA: SS-Gblock (10ng/uL)

*resuspend primers to 100uM

*dilute primers from 100uM to 10uM

2.5uL of 100uM to 10uM

22.5uL of nf water each.

PCR components:

98°C - 30s

10 cycles

98°C - 15s

60.0°C - 30s

72°C - 15s

20 cycles

98°C - 15s

71.0°C - 30s

72°C - 15s

72°C - 5min

4°C - inf.

NANODROP RESULTS

Table1

	Plasmid	Concentration	260/280	260/230
1	NapA-saCas9-His6 1	378.5 ng/uL	1.86	2.05
2	NapA-saCas9-His6 2	304.0 ng/uL	1.84	1.79
3	NapA-saCas9-His6 3	293.6 ng/uL	1.87	2.26
4	YcdB-saCas9-His6 1	282.5 ng/uL	1.88	2.28
5	YcdB-saCas9-His6 2	380.0 ng/uL	1.87	2.23
6	YcdB-saCas9-His9 3	303.3 ng/uL	1.84	1.81