Chimeric Transcription Factor Assay Protocol

Material

Plasmids used. The schematic maps are shown above.

pCAG- relA/NLS/traR-polyA

(trabox)₇-CMVmin-AtIPT4-IVS-IRES-LOG(pIRESneo3)

Enzymes to linearize the plasmids for electroporation

pCAG- relA/NLS/traR-polyA was cut by HindIII.

(trabox)₇-CMVmin-AtIPT4-IVS-IRES-LOG(pIRESneo3) was cut by *PvuI*.

Medium

DMEM 10% FBS

DMEM 10% FBS G418 (400 μ g/mL)

The primers for quantitative RT-PCR

• atIPT4

Forward: 5'- gtgcaacgacaaaatggtgg-3'

Reverse Sequence: 5'-gctaaccagcactagaagtcc -3'

• log1

Forward: 5'-ggactgatctctcaggctgtg-3'

Reverse: 5'-cgactacgtatagacgatggc-3'

Assay protocol

- 1, The EA.hy 926 cells were cultured to about $1.0 * 10^7$ cells/dish (the dish size is 10 cm in diameter) and used for electroporation.
- 2, C8 was added to the dish at final concentration of 0, 20, or 40 μ M and incubated further for 24 hours.
- 3, After harvesting the cells, total RNAs were purified according to the ordinary AGPC (Acid guanidinium thiocyanate-phenol-chloroform) method, and cDNAs were obtained by reverse transcription.
- 4, qPCR was performed using the above cDNA.

[note]

AGPC methods

- ①Vortex the collected cells in the tube for 10 to 15 sec.
- ②Add 1/10 volume of Na acetate.
- ③Add 1.4-times volume of Phenol/Chloroform/Isoamyl alcohol.
- 4 Vortex every 10 minutes.
- \bigcirc Centrifuge at 15000 rpm, 10 min, 4 $^{\circ}$ C.
- 6Recover the aqueous layer and mix with 400 $\,\mu$ L isopropanol.
- 7Leave at -20 ° C for 1 hour.
- & Centrifuge at 15000 rpm, 10 min, 4 \degree C and remove supernatant.
- \bigcirc Add 400 μ L of 75% ethanol, centrifuge at 15000 rpm, 10 min, 4 $^{\circ}$ C and remove supernatant.
- ① Dry the pellet and dissolve with 10 μ L of pH = 7.4 TE buffer

Reverse transcription

Total RNA	$8\mu\mathrm{L}$
Oligo (dT)[10 pmol/ μ L]	$1 \mu L$
5* RT buffer	$4 \mu L$
10mM dNTP	$2 \mu L$
ReverTra Ace	$1 \mu L$
RNase inhibitor	$1 \mu L$
dH2O	$3 \mu L$
total	$20~\mu\mathrm{L}$

After mixing, samples were heated at 42° C. for 20 min, 99° C. for 5 min, 4° C. for 5 min, then stored at -20° C.

·qF	CR
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2*SYBR	$10\mu\mathrm{L}$
10 mM Primer F	$0.4~\mu\mathrm{L}$
10 mM Primer R	$0.4~\mu\mathrm{L}$
dH2O	$6.2~\mu\mathrm{L}$
Sample	$3 \mu L$
total	20 μL

XSYBR: KAPA SYBR FAST qPCR kit