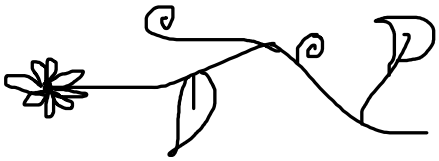
6/01/17

Thursday, June 1, 2017 9:04 AM



Who was in lab today: Jeff, Ayesha, Qingxi, Ana, Salma, Martin

The transformation performed on 5/31/2017 did not show expected results No colonies grew overnight.

Today we made two 500mL bottles of LB agar media.
See making LB media protocol.

IPTG--> isopropyl thiogalactoside

Plasmid amount added	Plasmid type	Cell culture type	Cell culture name	Type of agar plates used	PLATE #	TUBE NUMBER
25 uL		New	DH5alpha	New plate	1	1
25 uL		New	DH5alpha	Old plate	2	1
10uL		Old	DH5alpha	New plate	3	2
10uL		Old	DH5alpha	Old plate	4	2
25 uL		New	DH5alpha	New	5	3
25 uL		New	DH5alpha	Old	6	3
10 uL		Old	DH5alpha	New	7	4
10 uL		Old	DH5alpha	Old	8	4
40 uL		New	BL21 (DE3)	New	9	5
40 uL		New	BL21 (DE3)	Old	10	5
40uL		Old	BL21 (DE3)	New	11	6
40uL		Old	BL21 (DE3)	Old	12	6
25 uL	PET28B	New	DH5alpha	New	13	7
25 uL	PET28B	New	DH5alpha	Old	14	7
10uL	PET28B	Old	DH5alpha	New	15	8
10uL	PET28B	Old	DH5alpha	Old	16	8

125 uL of 100mM CaCl2
1 uL of DNA (we don't really take this into account when we wannt to calculate the X uL of cells and the sterile water that we want to add)
XuL of cells
total volume should be 250 uL with sterile water

Antibiotics added to the LB agar media and 1000X more concentrated than we want. In 500 mL of LB agar media, we added 100uL of CAM (chloromphenicol).

Working conc. of CAM is 33mg/mL.
33mg/mL *10mL = 330 mg

We added 400 uL (1/1000th dilution) of IPTG + Kan to LB+Agar

Red marked plates--> Kanamyacin
Brown--> IPTG
Black--> Chloromphenicol