

## OneTaq (NEB)

### Reaction setup:

We recommend assembling all reaction components on ice and quickly transferring the reactions to a thermocycler preheated to the denaturation temperature (94°C).

Add to a sterile thin-walled PCR tube:

Component	50 $\mu$ l (our lab)	50 $\mu$ l reaction	Final Concentration
5X <i>OneTaq</i> Standard Reaction Buffer*	10 $\mu$ l	10 $\mu$ l	1X
10 mM dNTPs ( <a href="#">#N0447</a> )	1 $\mu$ l	1 $\mu$ l	200 $\mu$ M
10 $\mu$ M Forward Primer	2 $\mu$ l	1 $\mu$ l	0.2 $\mu$ M
10 $\mu$ M Reverse Primer	2 $\mu$ l	1 $\mu$ l	0.2 $\mu$ M
<i>OneTaq</i> DNA Polymerase	0.125 $\mu$ l	0.25 $\mu$ l	1.25 units/50 $\mu$ l PCR**
Bodo	0.125 $\mu$ l		
Template DNA	variable	variable	< 1,000 ng
Nuclease-free water	to 50 $\mu$ l	to 50 $\mu$ l	

\**OneTaq* GC Reaction Buffer and High GC Enhancer can be used for difficult amplicons

\*\*For amplicons between 3–6 kb, use 2.5–5 units/50  $\mu$ l rxn

Notes: Gently mix the reaction. Collect all liquid to the bottom of the tube by a quick spin if necessary. Overlay the sample with mineral oil if using a PCR machine without a heated lid.

Transfer PCR tubes to a PCR machine and begin thermocycling:

Thermocycling conditions for a routine PCR:

STEP	TEMP	TIME
Initial Denaturation	94°C	30 seconds
30 Cycles	94°C	15-30 seconds
	45-68°C	15-60 seconds
	68°C	1 minute per kb
Final Extension	68°C	5 minutes

Hold	4-10°C	
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