OneTaq (NEB)

Reaction setup:

We recommend assembling all reaction components on ice and quickly transferring the reactions to a thermocycler preheated to the denaturation temperature (94°C).

Add to a sterile thin-walled PCR tube:

Component	<mark>50 μl (our lab)</mark>	50 µl reaction	Final Concentration
5X One <i>Taq</i> Standard Reaction Buffer*	<mark>10 µl</mark>	10 μl	1X
10 mM dNTPs (<u>#N0447</u>)	<mark>1 μΙ</mark>	1 μΙ	200 μM
10 μM Forward Primer	<mark>2 μl</mark>	1 μΙ	0.2 μΜ
10 μM Reverse Primer	<mark>2 μl</mark>	1 μΙ	0.2 μM
One <i>Taq</i> DNA Polymerase	<mark>0.125 μl</mark>	0.25 μl	1.25 units/50 μl PCR**
Bodo	<mark>0.125 μl</mark>		
Template DNA	variable	variable	< 1,000 ng
Nuclease-free water	<mark>to 50 μl</mark>	to 50 μl	

*OneTaq GC Reaction Buffer and High GC Enhancer can be used for difficult amplicons

**For amplicons between 3–6 kb, use 2.5–5 units/50 μl rxn

Notes: Gently mix the reaction. Collect all liquid to the bottom of the tube by a quick spin if necessary. Overlay the sample with mineral oil if using a PCR machine without a heated lid.

Transfer PCR tubes to a PCR machine and begin thermocycling:

Thermocycling conditions for a routine PCR:

STEP	TEMP	TIME
Initial Denaturation	94°C	30 seconds
30 Cycles	94°C	15-30 seconds
	45-68°C	15-60 seconds
	68°C	1 minute per kb
Final Extension	68°C	5 minutes

Hold	4-10°C	